

# **Cling-*E. coli* :**

## **Bacteria on target**



**Harvard iGEM 2007**

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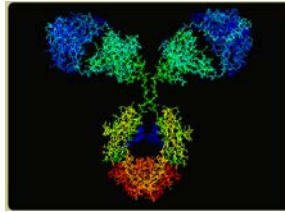
# The motivation

To develop a system for targeting bacteria to a specific substrate and effecting a cellular response

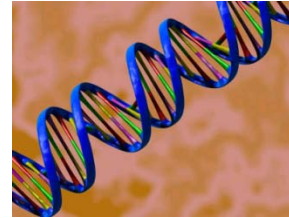


# Potential Targets and Applications

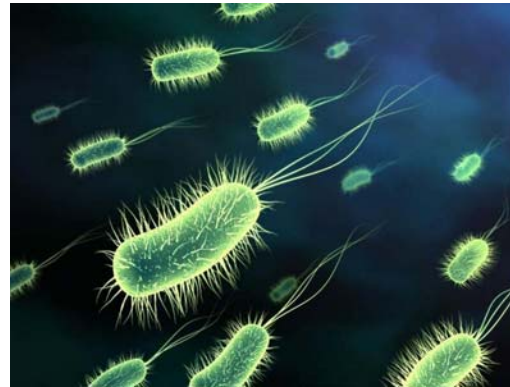
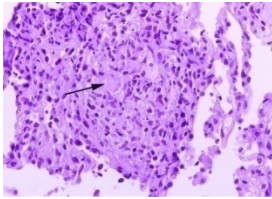
## Bind Proteins



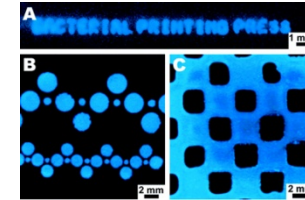
## Bind DNA/RNA



## Bind Tissue



## Bind Surface



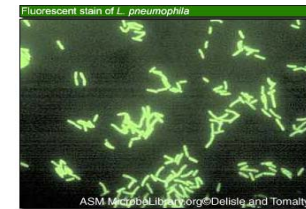
## Bind Viruses

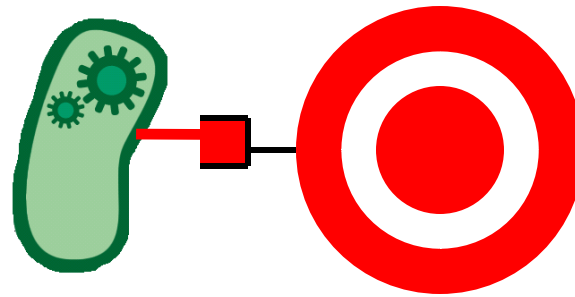


## Bind Drugs/Toxins

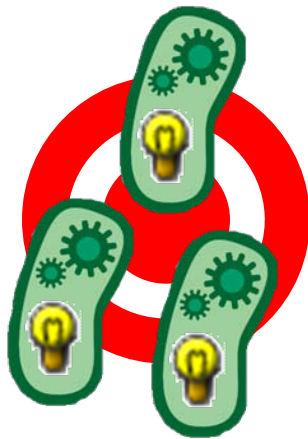


## Bind Other Cells

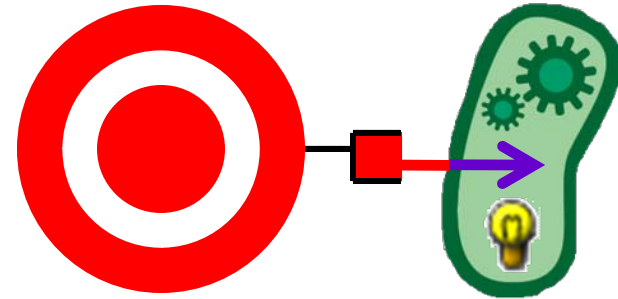




Bacterial targeting



Quorum-sensing

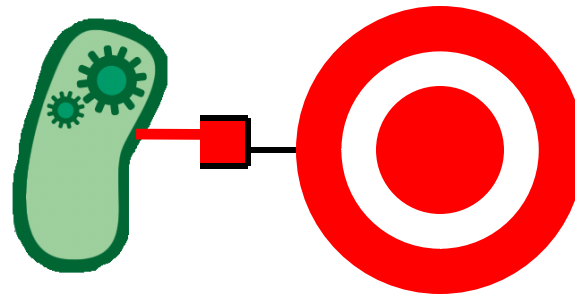


Fec signal transduction

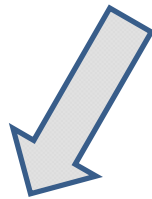
Introduction

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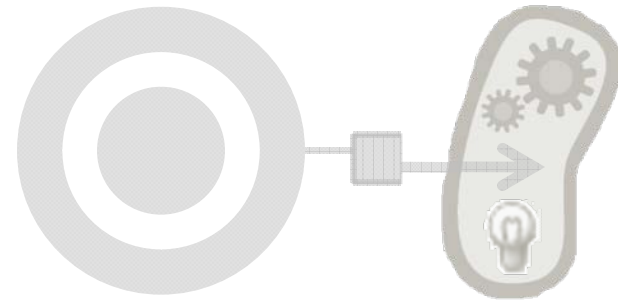
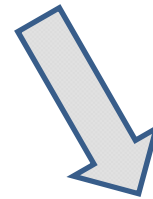




Bacterial targeting



Quorum-sensing

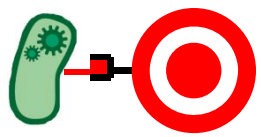


Fec signal transduction

Introduction

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# Surface-engineered bacteria

OmpA – C terminal insertion



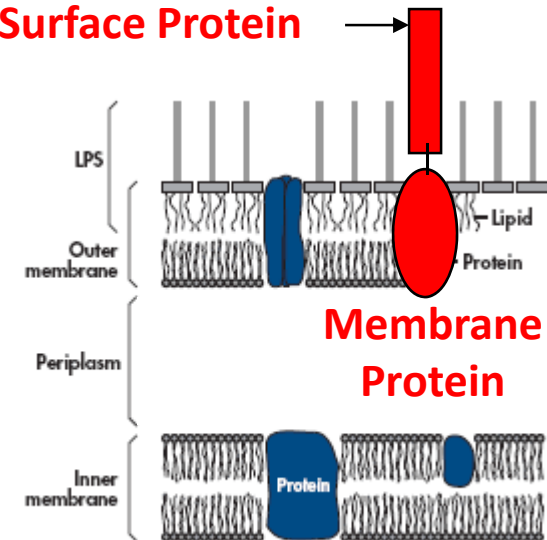
OmpA-Loop1 insertion

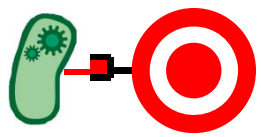


AIDA-1 – N terminal insertion



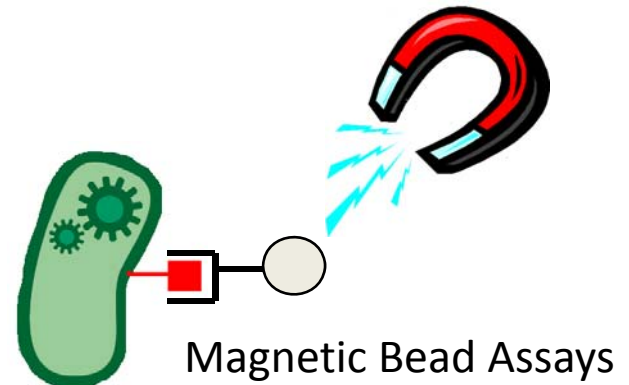
Surface Protein

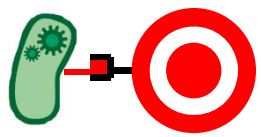




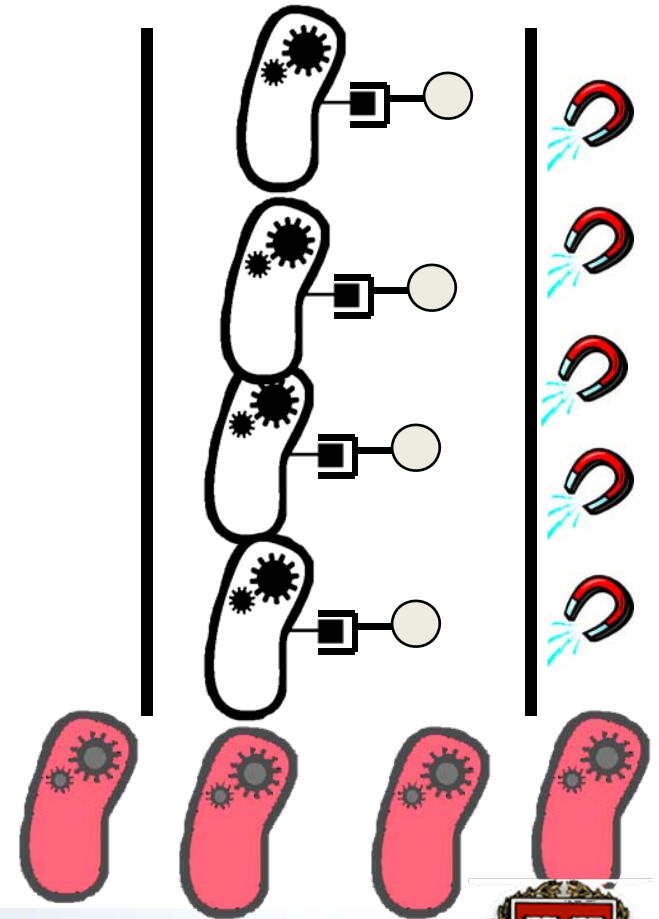
# Selecting/enriching for surface-engineered bacteria

- Tags
  - Histidine tag + nickel
  - Strep2 tag + streptavidin
- Assay
  - Magnetic Activated Cell Sorting (MACS)





# His and Strep2 tagged bacteria bind to beads

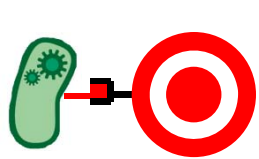


Bacterial Targeting

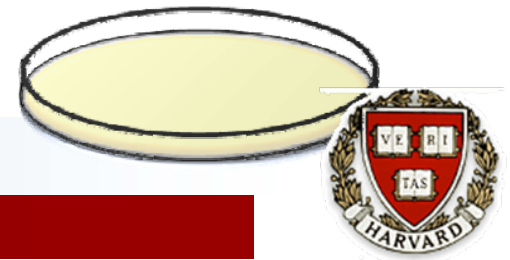
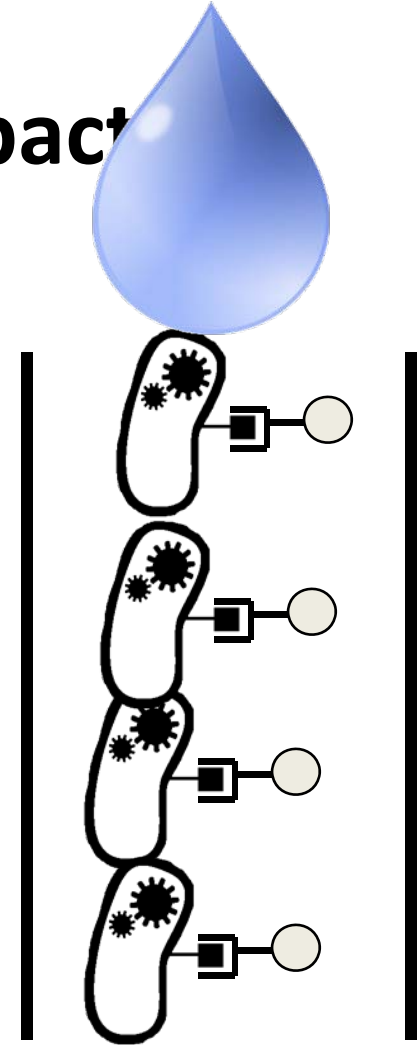
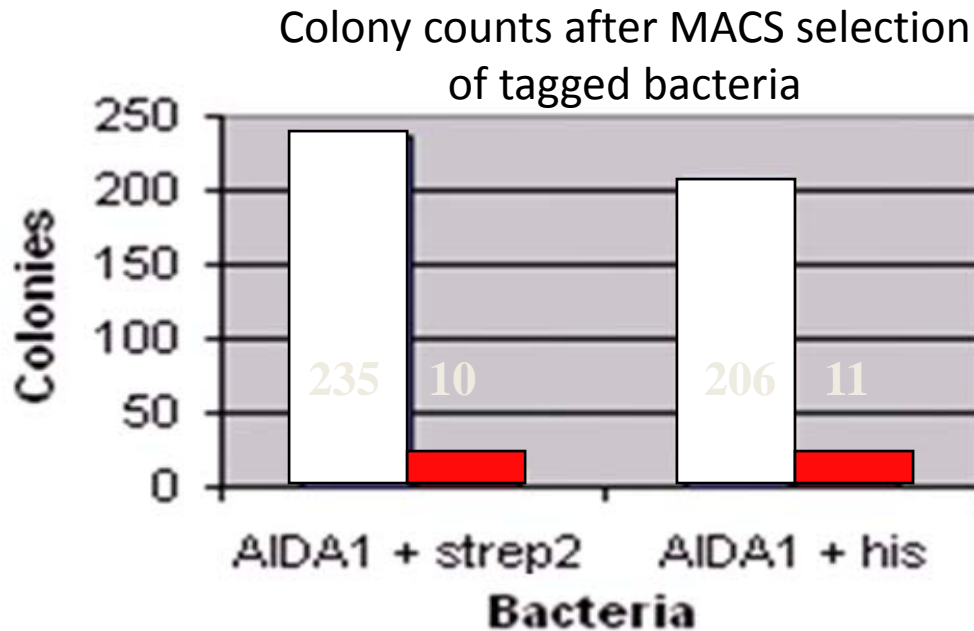
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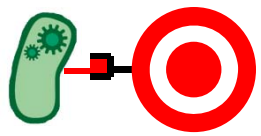






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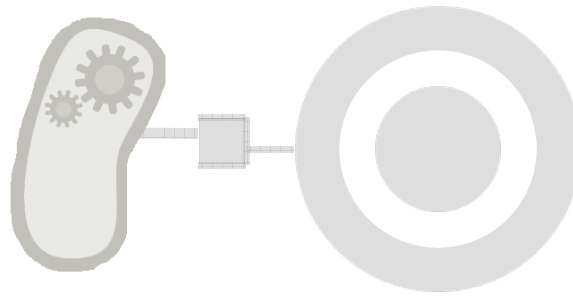




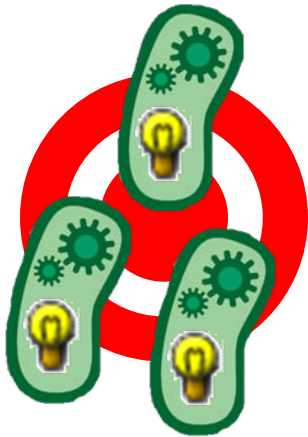
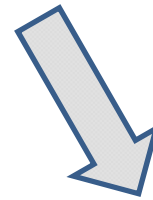
# Results

- Surface-expression vehicle – AIDA1
- Engineered surface-displayed histidine tag and strep2 tag
- Demonstrated bacterial targeting to nickel and streptavidin beads through MACS

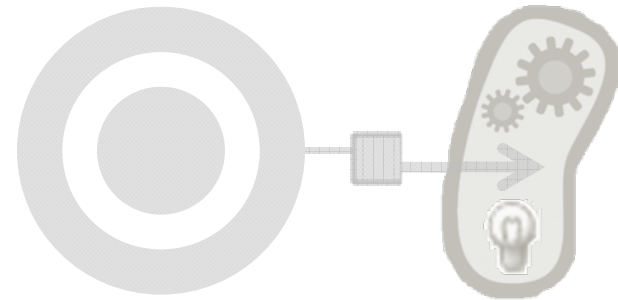




Bacterial targeting



Quorum-sensing



Fec signal transduction

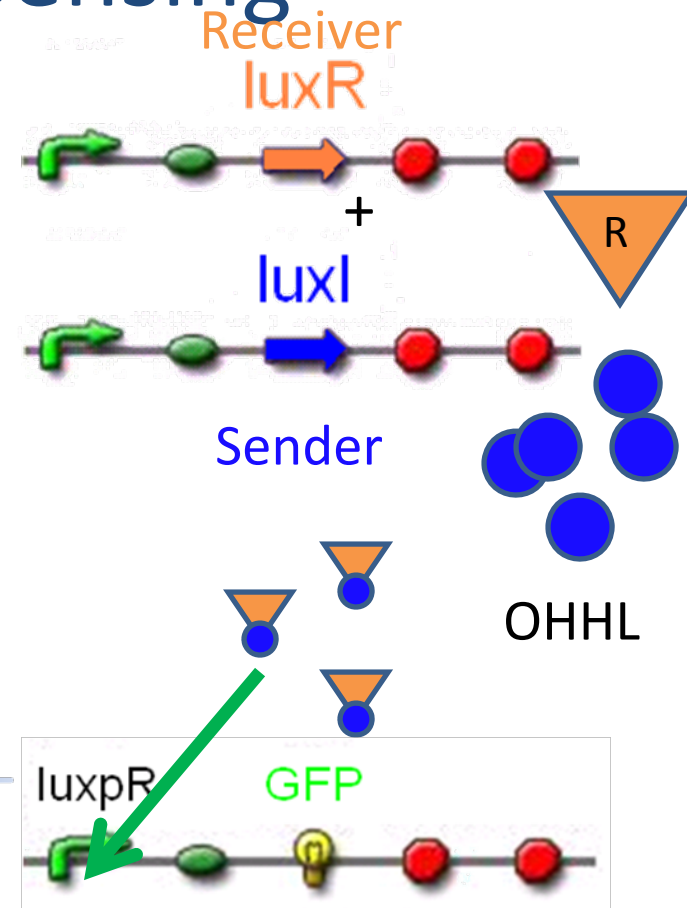
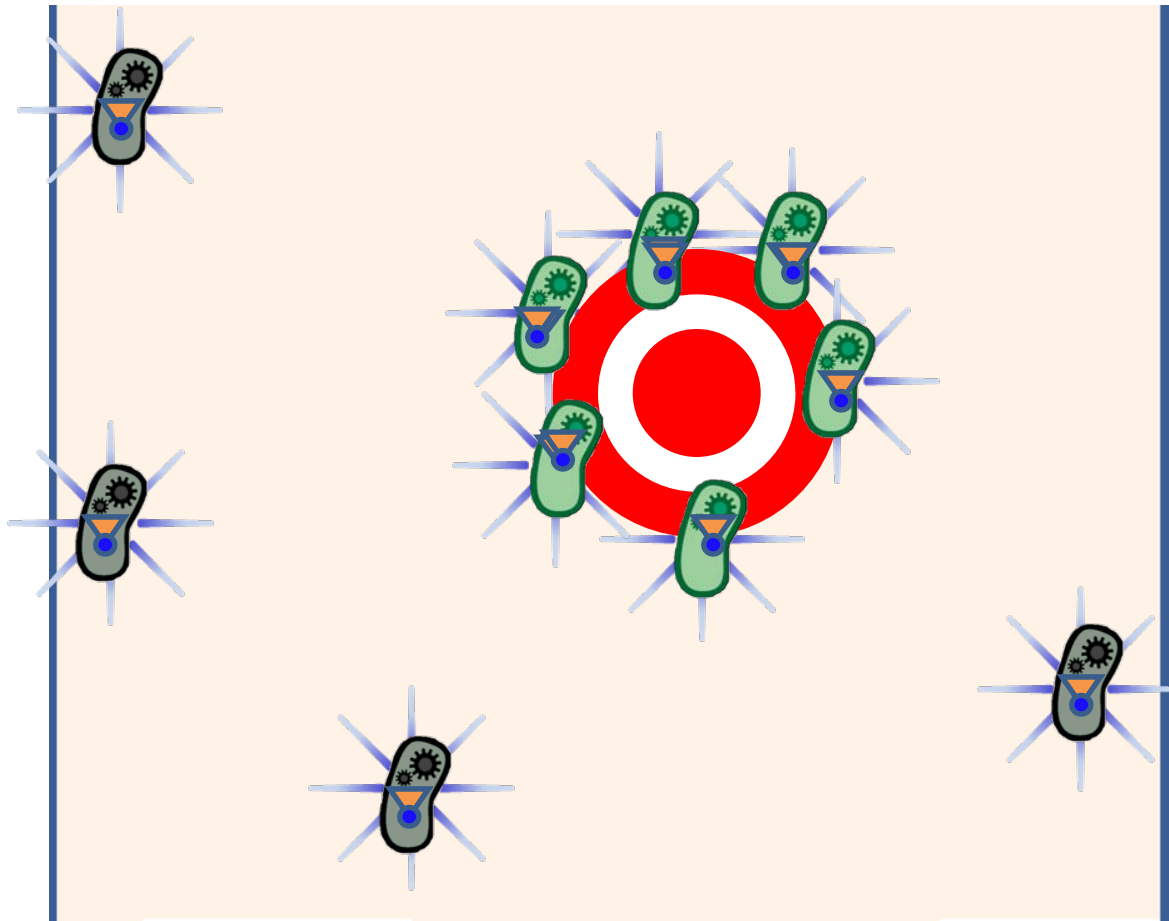
Quorum Sensing

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# luxI/luxR Quorum Sensing



Quorum Sensing

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# Cell-Cell Signaling Constructs

- Receivers (luxR + Reporter)

- GFP Receivers

- tetR controlled (Bba\_T9002)
    - Quorum controlled (Bba\_R0062 + Bba\_C0261 + Bba\_E0240)

- mRFP Receivers

- tetR controlled (Bba\_F2620 + Bba\_I13507)
    - Quorum controlled (Bba\_R0062 + Bba\_C0261 + Bba\_I13507)

- mCherry Receivers (Bba\_F2620 + Bba\_J06702)

- Senders (bicistronic luxI + Reporter)

- mRFP Sender

- tetR controlled (Bba\_S03623 + Bba\_I13507)
    - lacI controlled (Bba\_S03608 + Bba\_I13507)
    - Quorum controlled (Bba\_R0062 + Bba\_A340620 + Bba\_I13507)

- GFP Sender

- tetR controlled (Bba\_S03623 + Bba\_E0240)
    - lacI controlled (Bba\_S03608 + Bba\_E0240)
    - Quorum controlled (Bba\_R0062 + Bba\_A340620 + Bba\_E0240)

- mCherry Sender

- tetR controlled (Bba\_S03623 + Bba\_J06702)

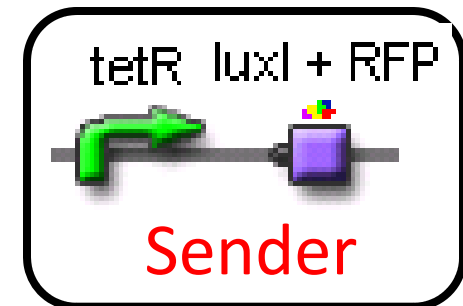
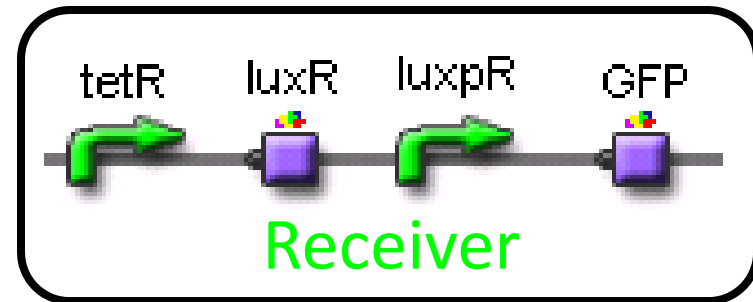
- Single Cell

- Constitutive (Bba\_J23039 + Bba\_T9002)
  - Quorum Controlled (Bba\_R0062 + Bba\_A340620 + Bba\_C0261 + Bba\_E0240)

- Construction Intermediates

## Quorum Sensing

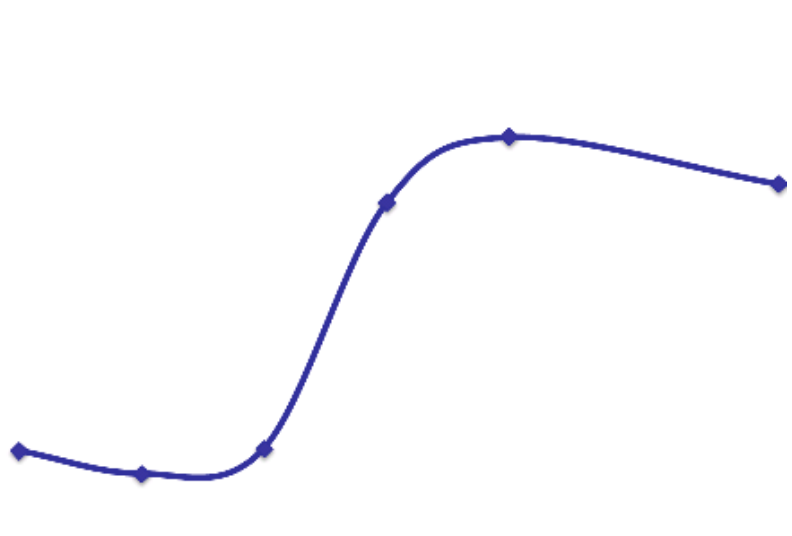
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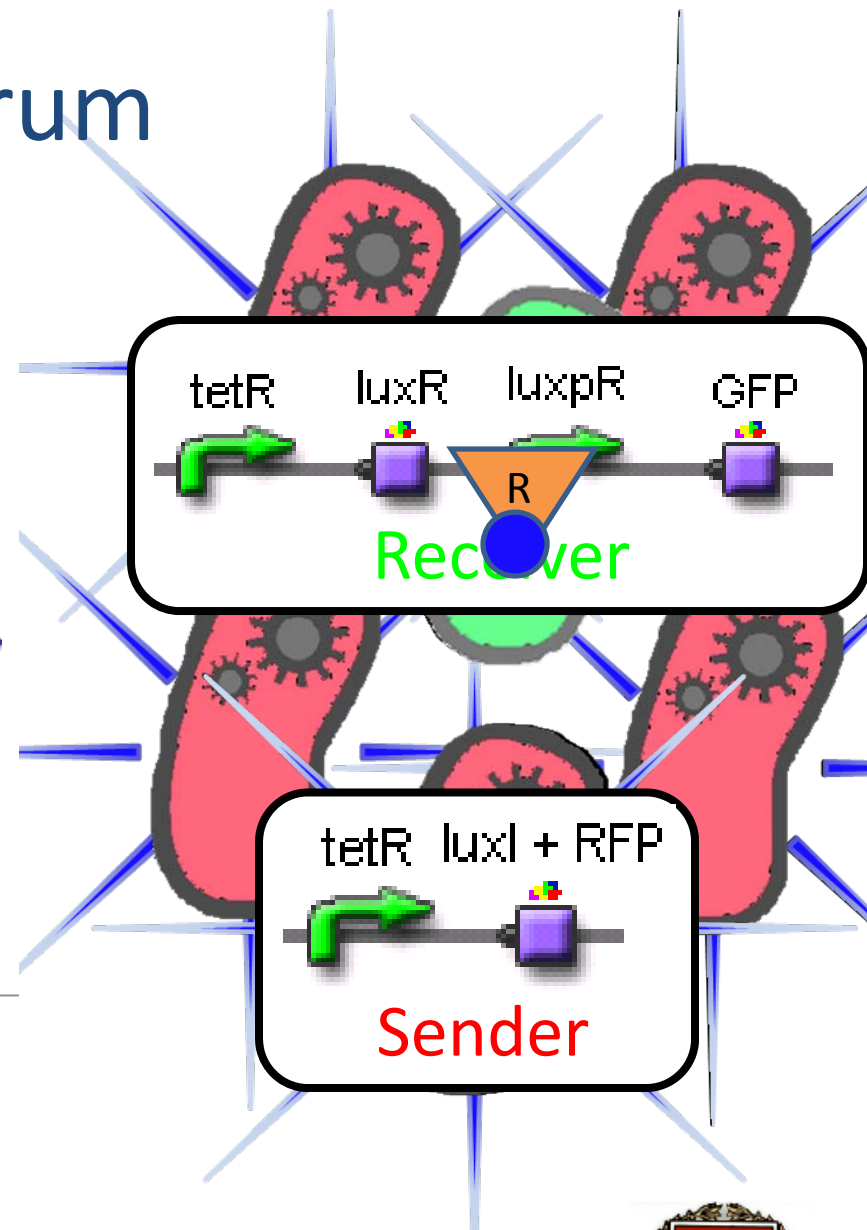


# Switch-like Quorum Response

GFP Fluorescence per OD  
(response of receiver)



Amount of Sender Cells Added



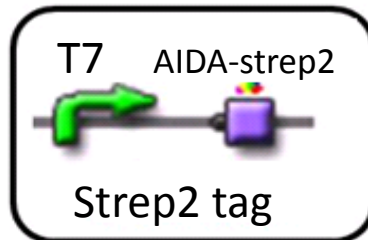
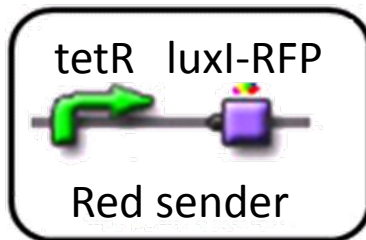
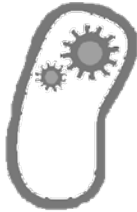
Quorum Sensing

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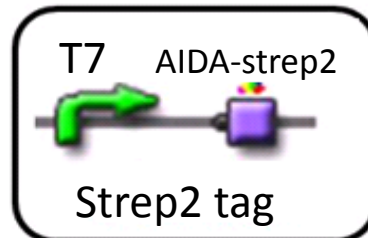
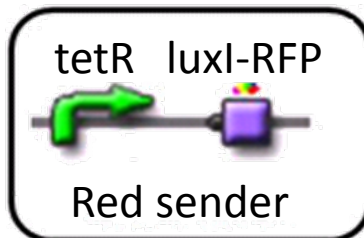
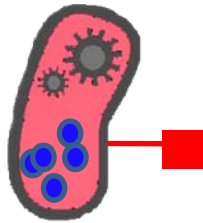


# MACS selection of cotransformed luxI-RFP/AIDA-strep2 cells





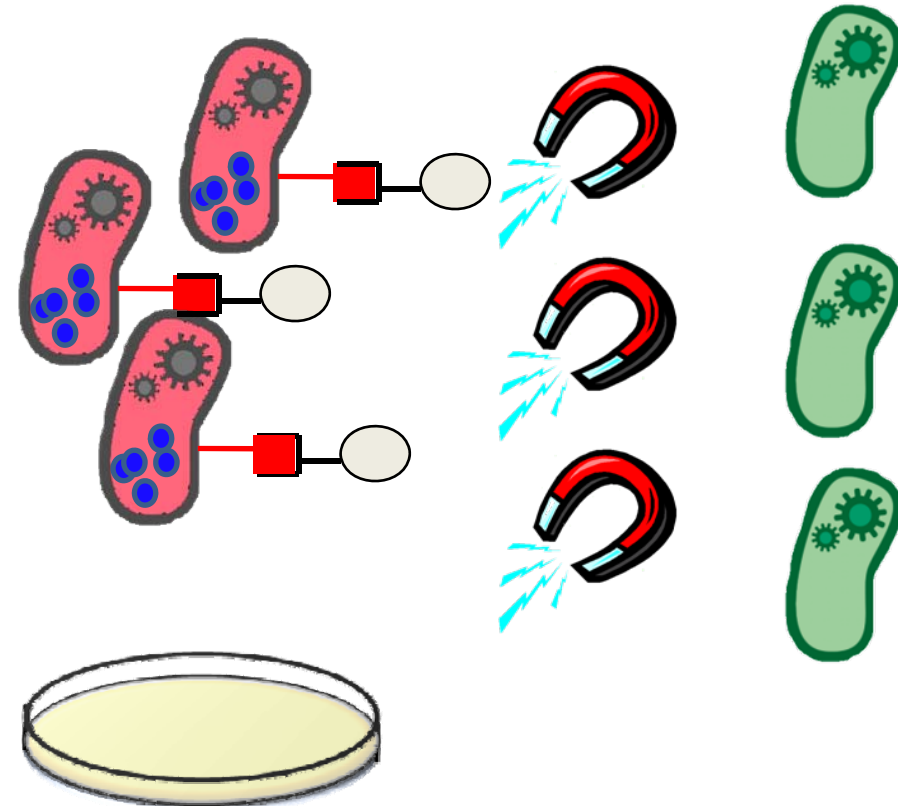
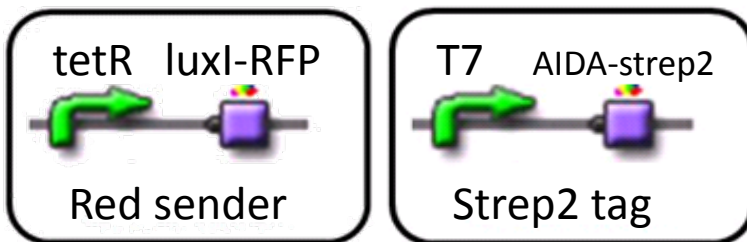
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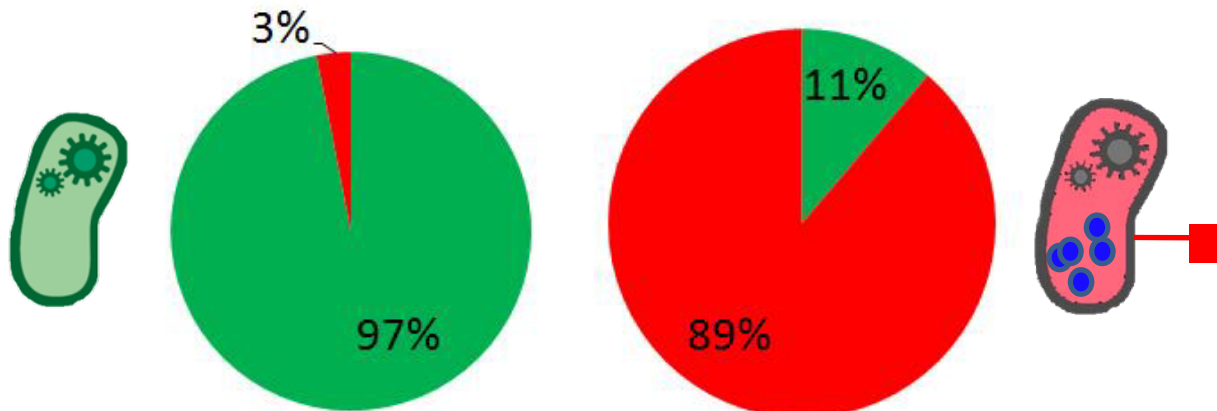
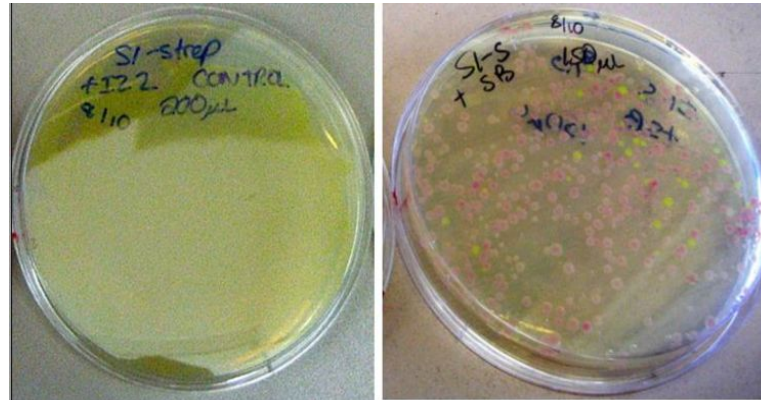




# MACS selection of cotransformed luxI-RFP/AIDA-strep2 cells

Before

After



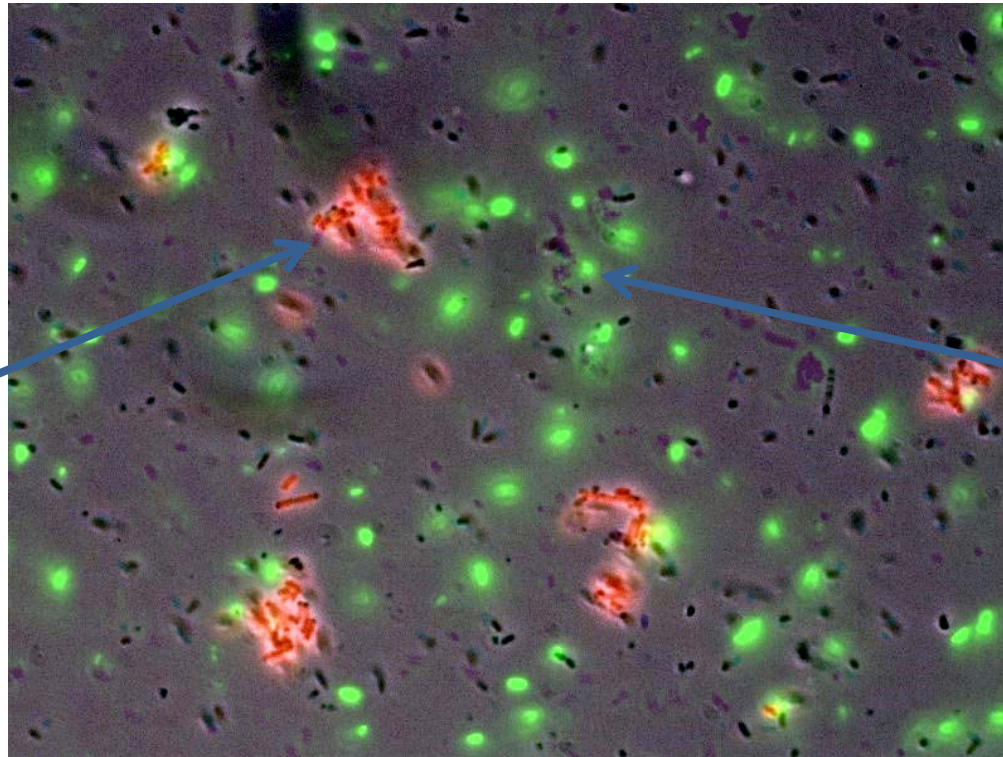
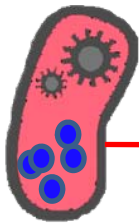
Quorum Sensing

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# luxI-RFP/AIDA-strep2 cells clump around streptavidin beads



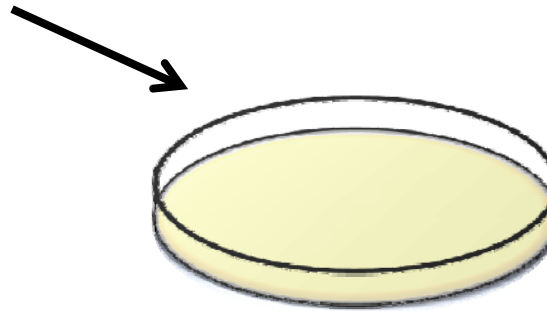
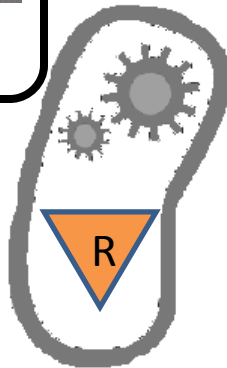
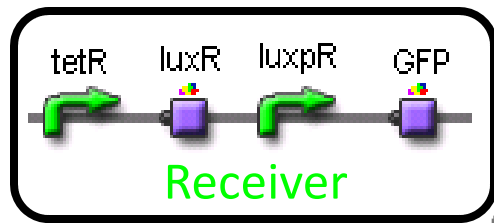
Quorum Sensing

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# Selected luxI-RFP/AIDA-strep2 cells can send quorum signal



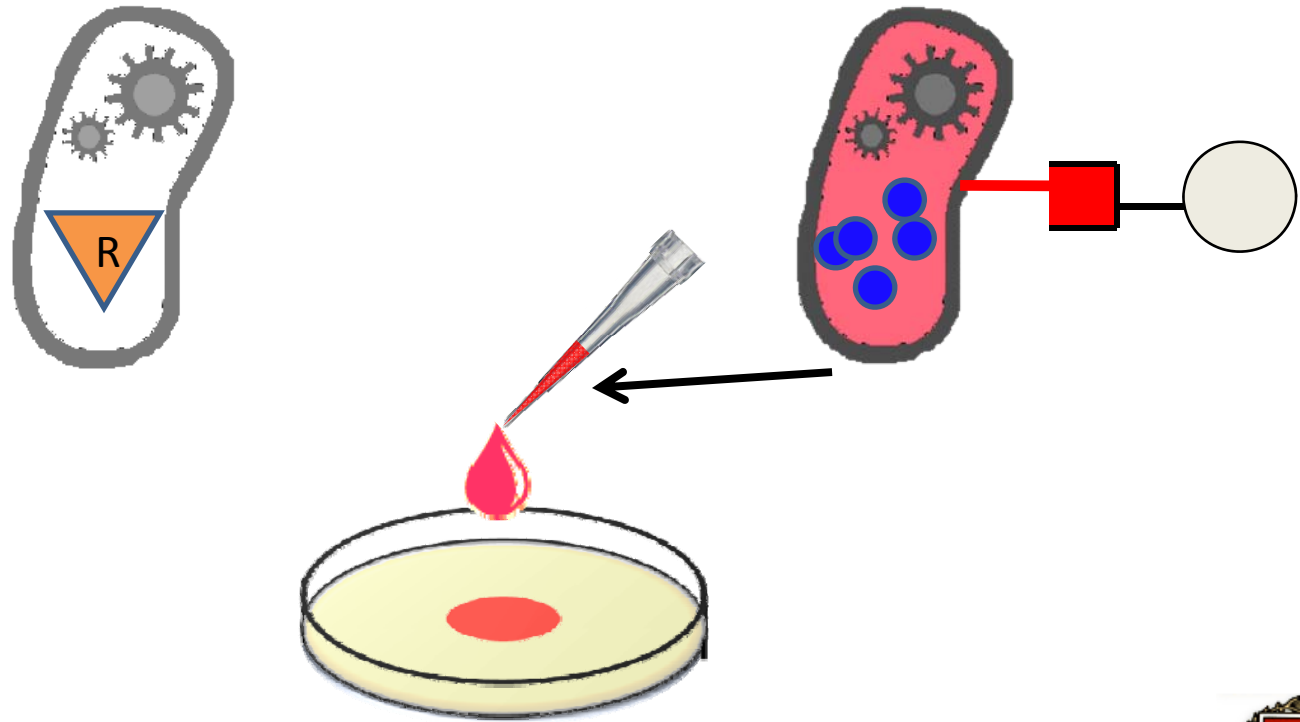
Quorum Sensing

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# Selected luxI-RFP/AIDA-strep2 cells can send quorum signal



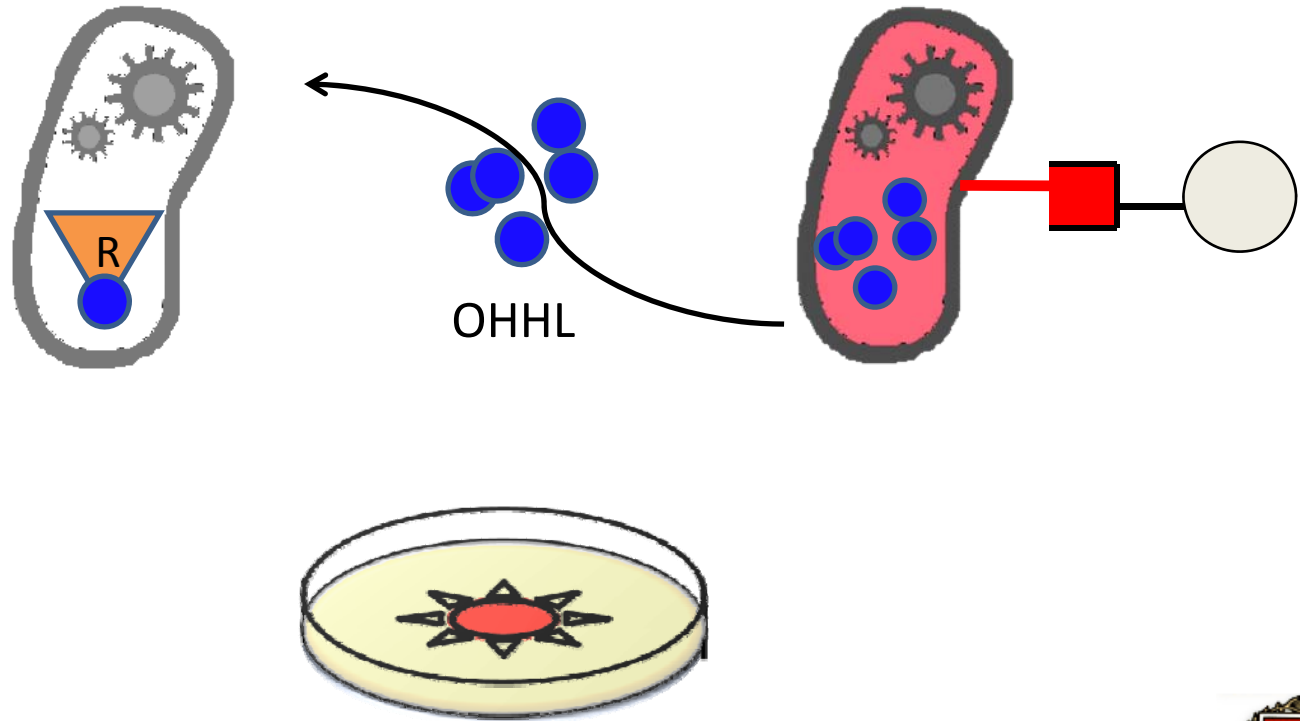
Quorum Sensing

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Quorum Sensing

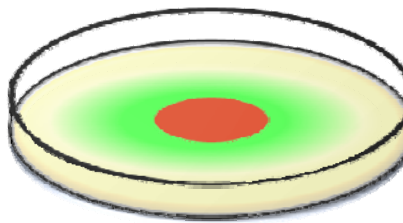
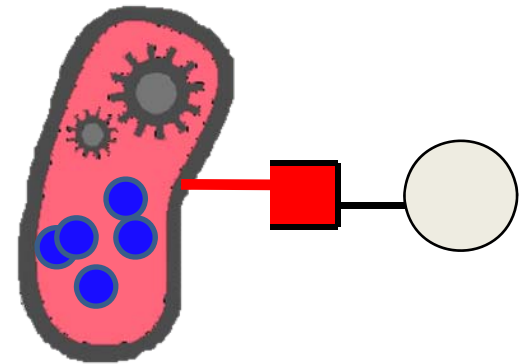
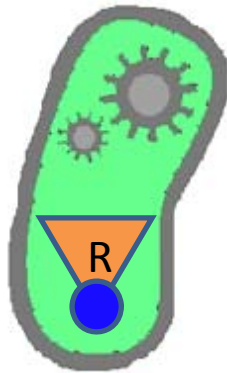
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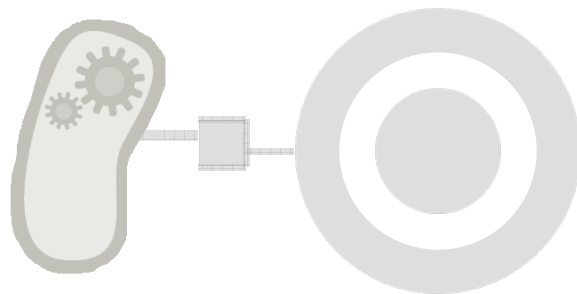
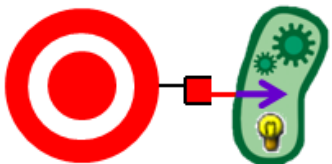
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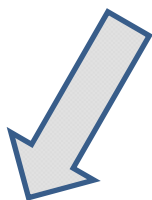
Quorum Sensing

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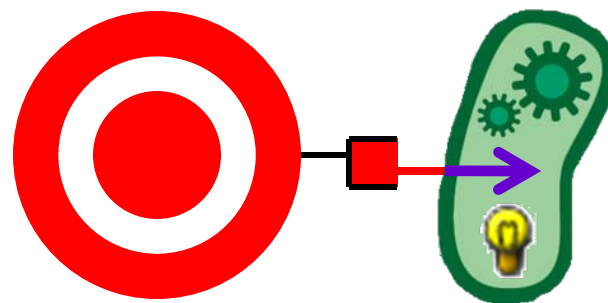




Bacterial targeting



Quorum-sensing



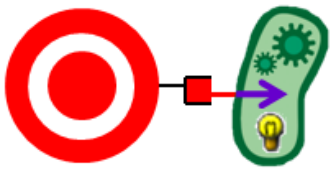
Fec signal transduction

Fec signal transduction

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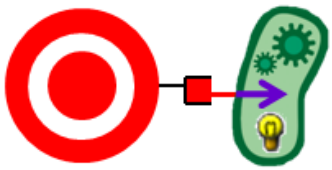




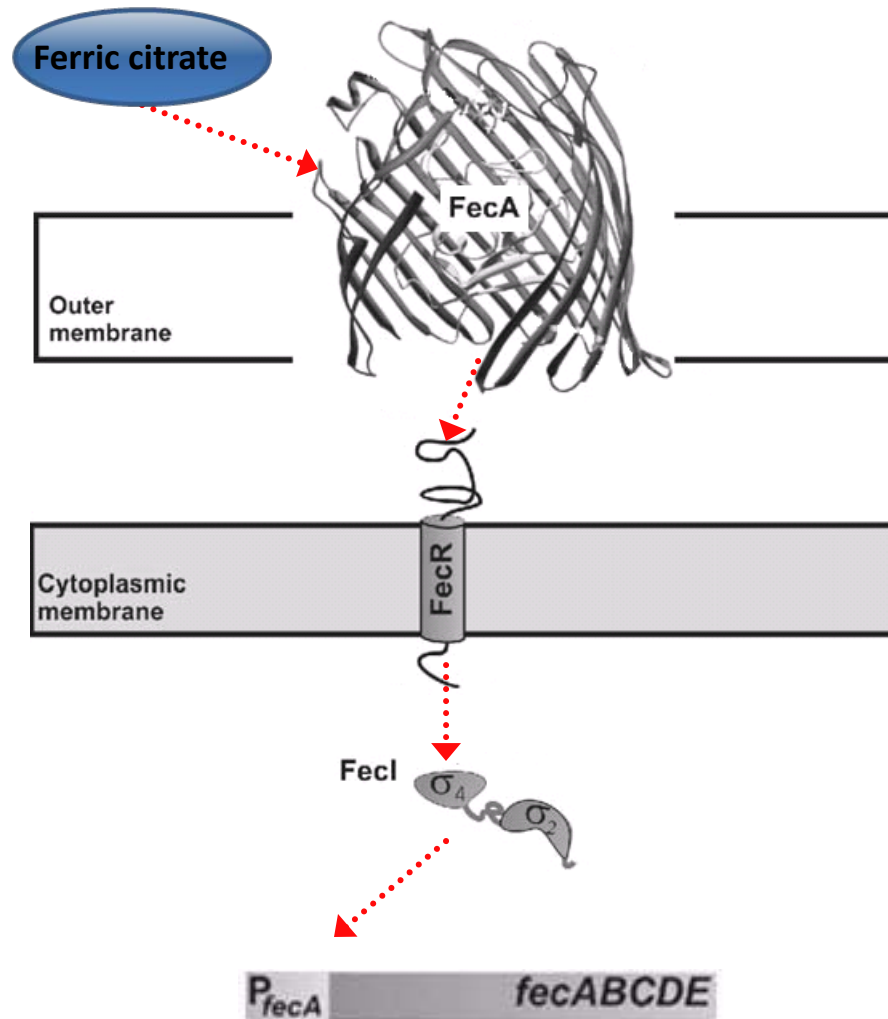
# Motivation: Fec System

- Goal: Direct cell signaling
- Method: Re-engineer an existing signal transduction pathway
- Fec system:
  - well-characterized
  - only outer membrane signal transducer





# Overview of Fec System

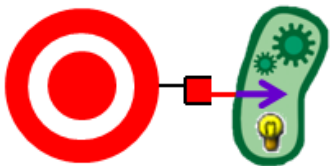


Fec signal transduction

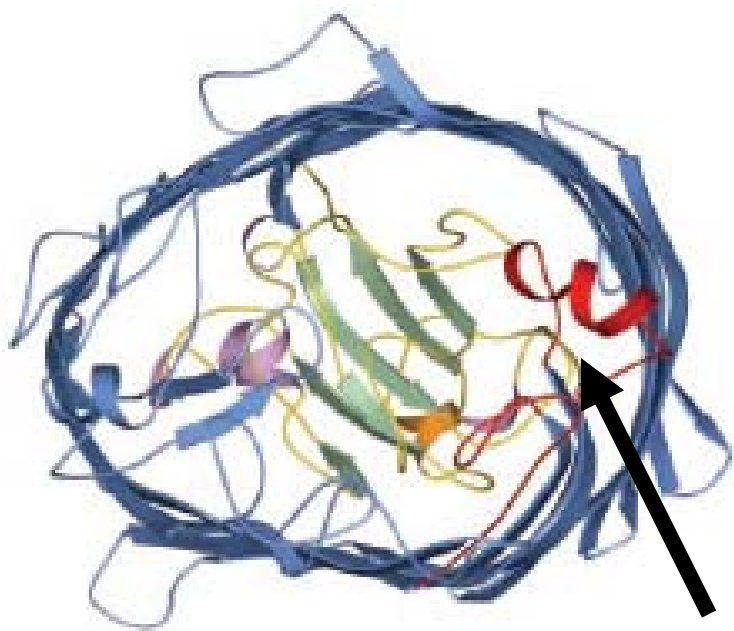
Harvard iGEM 2007

Braun et al. "Gene Regulation by Transmembrane Signaling."  
Biometals 2006 Apr;19(2):103-13.

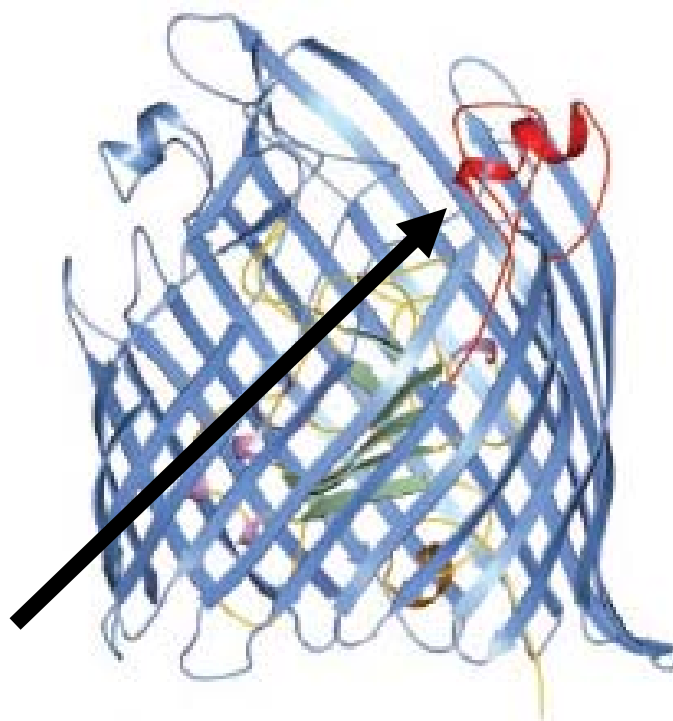




# Loops 7 and 8 as potential insertion sites



**Loops 7 & 8**

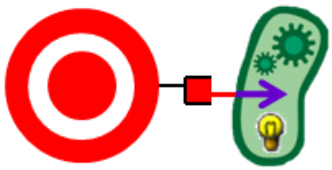


Fec signal transduction

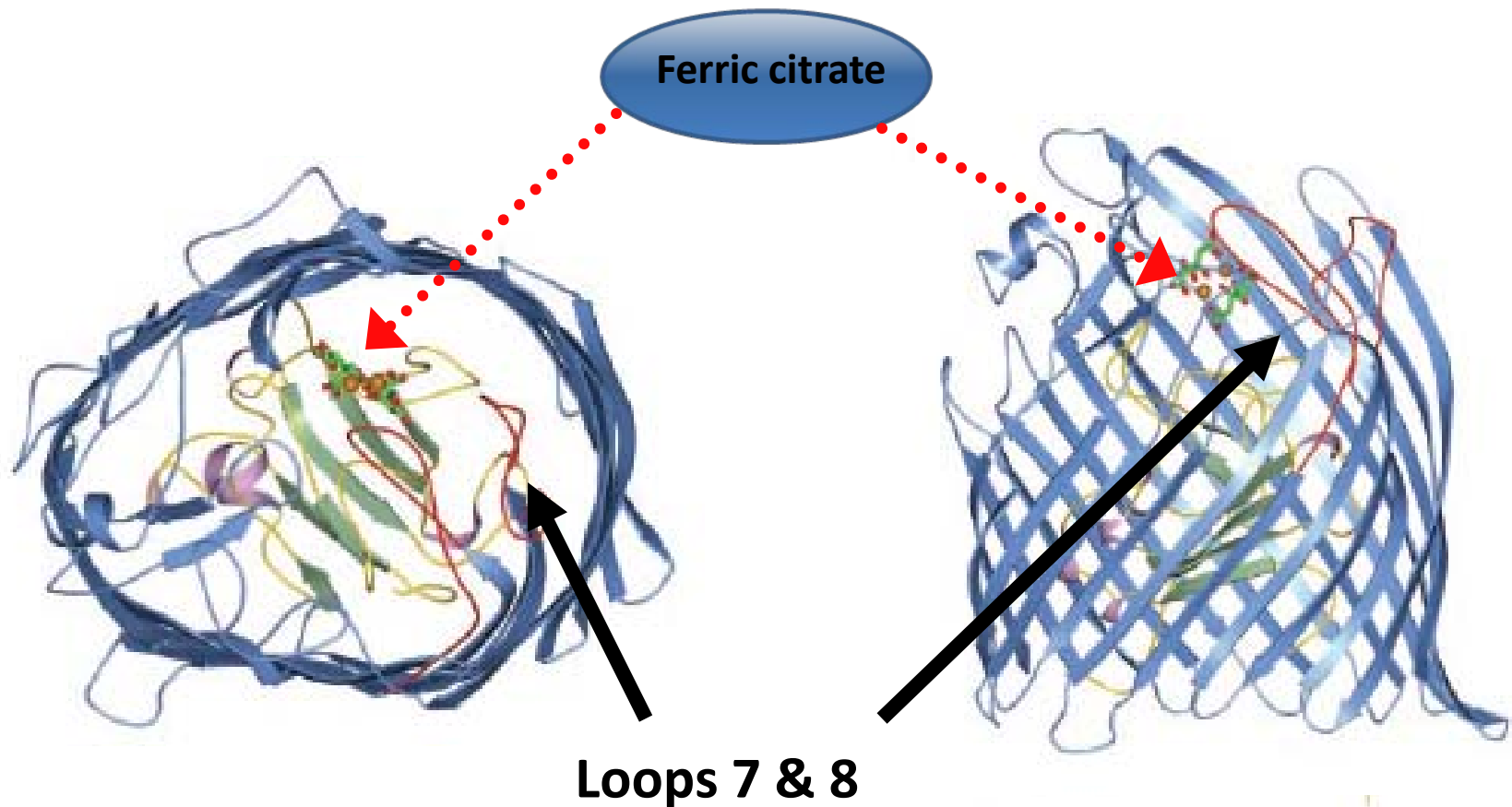
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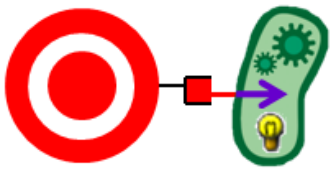
Ferguson AD, et al. Structural basis of gating by the outer membrane transporter FecA. Science 2002 Mar 1; 295(5560) 1715-9.





# Loops 7 and 8 as potential insertion sites

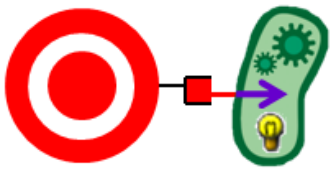




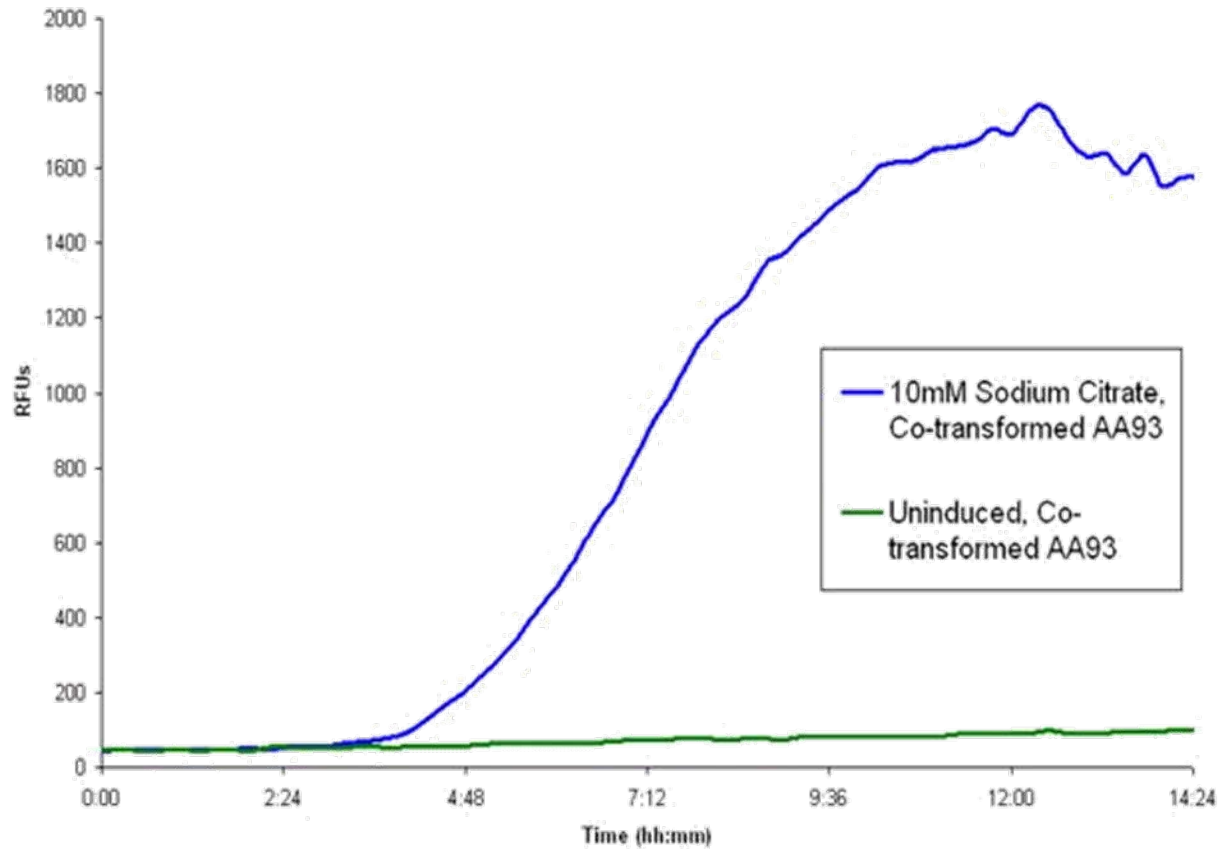
# Constructs

- From Braun lab (U. Tuebingen, Germany)
  - Fec knock-out strain, AA93
  - FecIRA plasmid
  - $P_{Fec}$ -GFP plasmid
- pColA Duet Vector
  - Allows regulated expression of Fec genes under T7 promoter





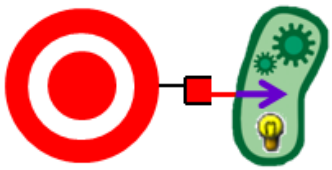
# Fec-induced GFP expression



Fec signal transduction

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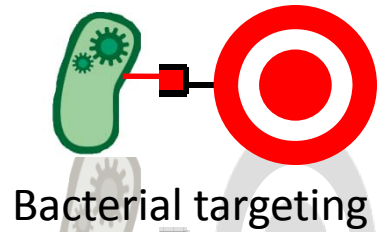


# Troubleshooting and Next Steps

- Problems:
  - Growth media: WL vs. LB?
  - Toxicity: membrane disruption?
- Goals:
  - Nickel and Streptavidin Binding
  - Finding new targets with signaling
    - Random library
    - Computational Approach



# Conclusions and Future Directions

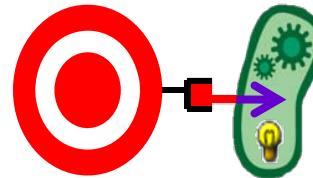
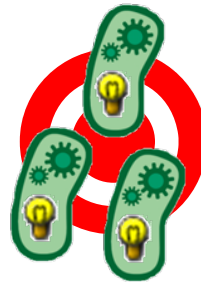


- Surface engineering – AIDA1
- Histidine/Strep2 tags
- MACS

*Random peptide libraries*

- One-cell/two-cell quorum systems
- Characterized
- **Targeted quorum senders**

*Optimize localized quorum response*



- Characterized wild-type Fec signaling

*Computational design*

Conclusion

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# Acknowledgements

## Advisors

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Tamara Brenner

## Special thanks to...

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Tuebingen)

Costas Maranas (Penn  
State U)

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Bill Senapedis

Mike Strong

Harris Wang

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Harvard Life Sciences Division

Harvard School of Engineering and  
Applied Sciences

Conclusion

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